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conceptual translation product (SEQ ID NO:2) of its longest ORF (GenBank accession number AF 237670). Underlined is a stop codon (TAA) at -6 to -4 that precedes the ATG in-frame. Numbers I-XI indicate kinase subdomains as defined by Hanks et al. (25), with invariant and nearly invariant amino acid residues highlighted in black and gray, respectively.--

[Page 6, lines 7-13, please replace the paragraph with the following paragraph:]

--Fig. 3: Amino acid alignments. (A) Alignment of putative kinase catalytic domain of SOS2 with *Saccharomyces cerevisiae* SNF1, SEQ ID NO:3, (23) and human AMPK kinases, SEQ ID NO:4 (24). Amino acid residues identical in at least two proteins are highlighted in black and conservative substitutions in gray. Mutations that abolish SOS2 autophosphorylation (see Fig. 4) are indicated; first \* is K40N,m and second is \* G197E, which corresponds to the *sos2-5 allele*. (B) Alignment of the C-terminal portion of SOS2 with the regulatory domains of *Schizosaccharomyces pombe* (yCHK1, SEQ ID NO:5) and human CHK1 (hCHK1, SEQ ID NO:6) kinases (27).--

Page 13, prenumbered lines 4-24, please replace the paragraph with the following paragraph:

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--Protein Expression. To produce bacterially expressed recombinant proteins, the coding region of SOS2, SOS2(K40N), and SOS2(G197E) cDNAs were amplified by PCR with primers harboring restriction sites, cloned in frame into *Bam*HI-*Eco*RI of pGEX-2TK (Amersham Pharmacia), and transformed into *Escherichia coli* BL21 DE3 cells (Amersham Pharmacia). Mutations K40N and G197E in the SOS2 protein were created by site-directed mutagenesis. For glutathione S-transferase (GST)-SOS2(K40N), primer pairs 5'-GCGGATCCATGACAAAGAAAATGAGAAGAGTGGGC (SEQ ID NO:7) and 5'-

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ATTGTA CTCTTAGCCATAATGTTGATGGCT (SEQ ID NO:8) were used for the first PCR, and 5'-GCGAATTCTTAAGTTGGGATCAAAACGTGATTGTTCTG (SEQ ID NO:9) and 5'-GTGATAATGTAGCCATCAACATTATGGCTA (SEQ ID NO:10) were used for the second PCR. For the mutant protein GST-SOS2 (G197E), primer pairs 5'-GCGGATCCATGACAAAGAAAATGAGAAGAGTGGGC (SEQ ID NO:11) and 5'-ATATAACGAAAAGAATAACCTCGCAAGACC (SEQ ID NO:12) were used for the first reaction and 5'-GCTGATATTTGGTCTTGCGAGGTTATTCTT (SEQ ID NO:13) and 5'-GCGAATTCTTAAGTTGGGATCAAAACGTGATTGTTCTG (SEQ ID NO:14) were used for the second reaction. The final amplification was done with 5'-GCGGATCCATGACAAAGAAAATGAGAAGAGTGGGC (SEQ ID NO:15) and 5'-GCGAATTCTTAAGTTGGGATCAAAACGTGATTGTTCTG (SEQ ID NO:16) on both templates. The final constructs were confirmed by sequencing *E. coli* cultures were induced with 0.5 mM isopropyl  $\beta$ -D-thiogalactoside, and recombinant proteins were affinity-purified from bacterial lysates with glutathione-Sepharose beads (Amersham Pharmacia).--

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Page 25 (Abstract), after the last line, beginning on a new page, please insert the attached Sequence Listing.

#### REMARKS

Claims 1-42 are pending in the present application.

Applicants have now submitted a Sequence Listing and a corresponding computer-readable Sequence Listing. The sequence information recorded in the corresponding computer-readable Sequence Listing is identical to the paper copy of the Sequence Listing. Support for all of the sequences listed in the Sequence Listing is found in the present